HisCube Ni-INDIGO His-tag MagBeads MINI Kit

Kit contents

1 x 5 ml PureCube Ni-INDIGO MagBeads
2 x 50 ml Binding Buffer
2 x 50 ml Wash Buffer
1 x 50 ml Elution Buffer
1 x Cube MagBead Separator
1 x 500 mM DTT (add 1,016 ml ddH2O)
1 x 500 mM EDTA (add 0.866 ml ddH2O)

Storage Temperature 4 °C / 39 °F

Protocol

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Fast Protocol

This „Fast Protocol“ is only meant as **reminder** for experienced HisCube Ni-INDIGO MINI Kit users

1. Pipet 200 µl Ni-INDIGO resin slurry into your reaction tube of 1.5 or 2 ml volume.
2. Equilibrate the MagBeads with 1.5 ml Binding Buffer by shaking the reaction tube for about **30 seconds** end-over-end. **Repeat** this step. Separate with the magnet and discard the supernatant.
3. Immediately before loading: Re-filter the cleared lysate through a 0.2 µm filter e.g. a syringe filter.
4. Load the required volume (max. 1.8 ml) of filtered cleared lysate. **Vortex for 15 sec**.
5. Repeat the vortexing of Step 7 every **15 minutes** for **1 hour** at **4 °C** or for **30 minutes** at room temperature.
6. Separate with the magnet for at least **20 sec**. Save the flow-through in a seperate tube and place the reaction tube containing the purification MagBeads back into the magnet separator.
7. Load the tube with up to 1.2 ml of Wash Buffer and separate for at least **20 sec**. Remove the supernatant. **Repeat** at least two times. Save all supernatants.
8. Elute the target protein by adding up to 100 µl Elution Buffer for **10 to 30 min** and then separate for at least **20 sec**. If necessary, repeat the elution step up to five times.

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